ADIAGENE
Bio-X Diagnostics

INSTRUCTION MANUAL

ADIAVET™ LEPTO REAL TIME

TEST FOR THE DETECTION OF *LEPTOSPIRA* BY REAL-TIME ENZYMATIC GENE AMPLIFICATION (PCR TEST)

Reference:
418026 (100 reactions)

VET

NOTE
Dye compounds in this product are sold under license from Biosearch Technologies, Inc. and protected by U.S. and worldwide patents either issued or in application. The license covers Veterinary Molecular Diagnostics.

English version
NE154-04
2020/01

ADIAVET™ LEPTO REAL TIME

Revision historic

N/A	Not Applicable (first publication)
Correction	Correction of document anomalies
Technical change	Addition, revision and/or removal of information related to the product
Administrative	Implementation of non-technical changes noticeable to the user

Note: minor typographical, grammar and formatting changes are not included in the revision historic

Release Date	Part Number	Change type	Change summaru
2014/09	NE154-01	N/A	First publication
2016/07	NE154-02	Administrative	Changing logos
2016/07	NE154-02	Administrative	Biosearch legal mention
2016/07	NE154-02	Administrative	Addition of table "Analysis options according to the specimen" §I.3.
2016/07	NE154-02	Correction	Modification of NucleoSpin Tissue protocol §IV.2.
2016/11	NE154-03	Technical change	Addition of swine and rodent samples
2020/01	NE154-04	Technical change	Addition of a NF-Water tube in the kit Modification of bood protocol (possibility to use sterile distilled water) Addition of water protocol

I. General informations

1. Purpose of the test

ADIAVET™ LEPTO REAL TIME kit is intended to detect pathogenic *Leptospira* using real-time Polymerase Chain Reaction (PCR) technology from swab, tissue, whole blood and urine specimens of rodent, bovine, ovine, caprine, equine, swine and dog and water.

2. Leptospirosis

Leptospirosis is a bacterial disease of world geographical distribution, affecting humans and animals. Generally, chronic infections are more frequent than the acute or subacute infections. In the large majority of the cases, the infections are asymptomatic. The chronic forms are dominated by abortions, interstitial nephritides and in certain species by infertility and uveitis. In the subacute forms, general symptoms are less intense and abortions frequent. Acute forms are characterized by septicaemias, hepatitises, nephritides and haemoglobinuries. Acute forms treatment must intervene in an early stage to be effective. Detection of pathogenic agent in biological samples (blood during septicaemia, tissues and urines for the chronic forms) allows an effective diagnostic approach. But direct observation is not very discriminating, isolation is difficult (sensitive germs).
Bacteriological diagnosis of Leptospirosis is thus not very operational. Leptospires are divided into two species according to serology: *Leptospira biflexa s.l.* gathering saprophyte *leptospira* and *Leptospira interrogans s.l.* including different serogroups of pathogenic *leptospira*. This last specie is currently divided into several genomic species.
The genetic classification, wich is based on DNA homology, divides leptospiral strains into four non pathogenic species: *L. biflexa, L. myeri, L. parva and L. wolbachii* and seven pathogenic species: *L. interrogans, L. borgpetersenii, L. weilii, L. nugochii, L. santarosai, L. inadai and L. kirschneri.*
The ADIAVET™ LEPTO REAL TIME PCR kit only detects the 7 pathogenic species.

3. Description and purpose of the test

This test is based on enzymatic gene amplification or PCR technology.
Amplified products are detected in real-time thanks to a specific labelled hydrolysis probe (5'-exonulease technology).

The ADIAVET™ LEPTO REAL TIME kit enables the simultaneous detection of:

- *Leptospira* (probe labelled in FAM),
- an internal control of amplification step specific from an exogenous DNA (probe labelled with a fluorochrome read in the same spectra as VIC and HEX).

Adiagene recommends using this test with DNA purification kits (Bio-X Diagnostics, Qiagen and Macherey-Nagel). Other purification kits can be used if they have been validated by the user.

Analysis options according to the specimen:

Specimen	Individual analysis
Swab (placental, vaginal…)	☑
Tissue (placenta, foetal tissues…)	☑
Whole blood	☑
Urine	☑
Water (environnemental)	☑

II. Material & reagents

1. Reagents provided with the kit

REF 418026 (100R)	
A5 ..... amplification solution	2 x 1000 µl tubes with green caps (a ready-to-use reagent)
LEPTO CTL+ ..... positive control *Leptospira*	1 tube with purple cap (to reconstitute)
NF-Water ..... Nuclease free Water	1 x 1000 µl tube with white cap (a ready-to-use reagent)

2. Validity and storage

On receipt, the kit should be stored at <**-15°C**.
It is recommended to make fractions of A5 solution if it should be defrosted more than 3 times.
Do not defrost reagents more than 3 times.
Realtime reagents are susceptible to light: store them in the darkness.
The A5 reagent is ready to use for PCR reaction.
Do not mix reagents of two different batches.

3. Use of LEPTO CTL+

LEPTO CTL+ is a positive control of amplification.
Add **200 µl** of **NF-Water** to the **LEPTO CTL+** tube and vortex at least 20 seconds.
Aliquot this solution by 6 or 12 µl and store them to <-15°C.
For each analysis, use **5 µl** of **LEPTO CTL+** in a well.

4. Equipment required but not supplied in the kit

Material should be Nuclease-free (e.g. autoclaved 25 minutes twice at +120°C or once 60 minutes at +121°C)

- Thermal cycler with consumables for real-time PCR: 0.2 ml PCR tubes or closed 96-wells PCR plates with optical quality.
- Class II Microbiological Safety Cabinet
- Centrifuge for microtubes, tubes of 5, 10 or 15 ml
- Etuve, heating baths or block heaters
- Vortex
- 1 - 10 µl pipette, 20 - 200 µl pipette and 200 - 1000 µl pipette
- Nuclease-free filter tips
- Nuclease-free microtubes: 1.5 ml and 2 ml
- Sterile tube of 5, 10 or 15 ml
- Latex or nitrile powder-free gloves
- Scalpel blades
- 96-100% ethanol solution
- Nuclease-free water
- PBS buffer
- Sterile distilled water

- DNA extraction kit (individual silica columns)

- QIAamp® DNA Mini Kit (Qiagen, 50 tests: ref. 51304 or 250 tests: ref. 51306)
For blood samples : Buffer EL (Qiagen, 1000 ml: ref. 79217)
- NucleoSpin® Tissue (Macherey-Nagel, 50 tests: ref.740952.50 or 250 tests: ref. 740952.250)

- Automated DNA/RNA extraction kit using magnetic beads

- ADIAMAG (Bio-X Diagnostics, 200 tests, ref.NADI003)

III. Recommendations before the analysis of samples

Before starting the test, read the entire protocol and scrupulously respect it.

1. Precautions

Adiagene has elaborated this PCR test with the use of Bio-X Diagnostics, Qiagen and Macherey-Nagel extraction kits. Other extraction kits can be used with a previous validation.

Follow the supplier's instructions for the storage, the preparation and the use of the extraction reagents.

Some kits include and/or need the use of toxic reagents. These reagents should be handled with gloves and into a chemical cabinet.

We strongly recommend that only appropriately trained personnel perform this test. Ensure the accuracy and precision of the micropipettes used. The quality of the obtained results depends upon rigorous respect of good laboratory practices.

The PCR generates large amount of amplified DNA. A few molecules of amplified products are sufficient to generate a positive result. It is important to reserve 2 rooms, one for manipulation of samples to be tested, and another one for amplified products analysis. Do not open the PCR tubes after amplification.
Samples for analysis should be handled and disposed of as biological waste. **Take all measures of security and confinement required for the manipulation of the concerned biological agents.**

We recommend using fractions of demineralised and saline water and to autoclave them 25 minutes at +120°C. Take a new fraction for each new manipulation to avoid contamination.

2. Storage of samples and DNA extracts

Samples can be stored a couple of days at +2/8°C. After 2 days, we recommend to store them at <-15°C.
Samples of blood with anticoagulant reagent must not be frozen.
Extracted DNAs are quite sensitive molecules. Crude extracts should be stored at the end of extraction on melting ice or at +2/8°C for 24 hours, then at <-15°C.

3. Samples preparation

See § IV for the extraction and purification of DNA.

4. Controls preparation

The use of controls allows verifying the reliability of the results.
The controls are included per trial of analysis. A trial is defined as all the samples treated in the same conditions.

The amplification step, for all the types of samples, is validated with the association of the controls included in the kit.

- The internal control included in A5 reagent verifies the amplification steps of each sample.
- The LEPTO CTL+ validates the amplification of the target.

Other controls must or could be added:

- **Negative control of extraction (required)**

To verify the absence of cross-contamination, at least one negative control must be included per trial. The control is a negative sample, for example a buffer used for dilutions.

- **Positive control of extraction (recommended)**

A positive control could be added in each trial. The control is a sample including *Leptospira*. It could come from a positive sample available in the laboratory or from a negative sample spiked with a solution of *Leptospira*. This positive control will be closed to the limit of detection of the method. It will inform about the fidelity of the obtained results between different trials.

IV. Extraction and Purification

1. Extraction using QIAamp® DNA Mini kit

All the centrifugations are performed at room temperature, except other indications.

Particular case of placentas:
May contain a large amount of microorganisms, manipulate them with extreme precaution.
1st method
Cut the cotyledon with a scalpel, then rub inside with a swab.
Perform analysis according to swab protocol.
2nd method
Perform analysis according to tissue protocol.

	Swab	Tissue	Bood	Urine / Water
Preparation of the sample	Mix the **swab** with **1 ml** of **1X PBS buffer.** Transfer **200 µl** in a microtube.	Put **20-30 mg** of **tissue** in a microtube.	Place **1 ml** of **blood** in a tube. Add **5 ml** of **EL buffer** or **sterile distilled water** Vortex, then incubate 15 minutes on melting ice (~0°C). Centrifuge 30 minutes at 10 000 g at 4°C. Discard the supernatant.	Place **10 ml** of **urine or water*** in a tube. Centrifuge 30 minutes at 10 000 g or 10 minutes 4 500 g Discard the supernatant.
Lysis	Add **180 µl** of **ATL buffer**, **20 µl** of **proteinase K**. Vortex. Incubate **30 minutes** at **+70°C** (or a night at +56°C).			
	Add **200 µl** of **AL buffer**. Vortex. Incubate **10 minutes** at **+70°C**.			
Binding preparation	Add **200 µl** of **ethanol 100%**. Homogenise the mixture by pipeting (~10 times) or by vortex (~15 secondes).			
Transfer to columns and binding to the membrane	Identify columns, apply the **whole** obtained solution to the corresponding column. Centrifuge 1 minute at 10 000 g. *If the whole sample has not been loaded once, apply the residual volume onto the column and centrifuge 1 minute at 10 000 g.*			
1st wash	Change the collection tube and add **500 µl** of **AW1 buffer** to the column. Centrifuge 1 minute at 10 000 g.			
2nd wash	Change the collection tube and add **500 µl** of **AW2 buffer** to the column. Centrifuge 1 minute at 10 000 g.			
Column dry step	Change the collection tube. Centrifuge 3 minutes at 10 000 g.			
Elution	Transfer the column to a microtube. Add **200 µl** of **AE buffer**. Incubate ~1 minute at room temperature and centrifuge 1 minute at 10 000 g.			
Storage	Close the tubes, identify and store at +2/8°C for 24 hours, then at <-15°C.			

*it's possible to centrifuge until 50 ml of water. In this case, after the discarding the supernatant is removed, add 1 ml of 1X PBS to the pellet, homogeneize and transfer 200 µl in the new microtube. And then, continue the protocol.

2. Extraction using NucleoSpin® Tissue kit

All the centrifugations are performed at room temperature, except other indications.

Particular case of placentas:
May contain a large amount of microorganisms, manipulate them with extreme precaution.
1st method
Cut the cotyledon with a scalpel, then rub inside with a swab.
Perform analysis according to swab protocol.
2nd method
Perform analysis according to tissue protocol.

	Swab	Tissue	Urine / Water
Preparation of the sample	Mix the **swab** with **1 ml** of **1X PBS buffer.** Transfer **200 µl** in a microtube.	Put **20-30mg** of **tissue** in a microtube.	Place **10 ml** of **urine** or **water*** in a tube. Centrifuge 30 minutes at 10 000 g or 10 minutes at 4 500 g. Discard the supernatant.
Lysis	Add **180 µl** of **T1 buffer**, **25 µl** of **proteinase K**. Vortex. Incubate **30 minutes** at **+70°C** (or a night at +56°C).		
	Add **200 µl** of **B3 buffer**. Vortex. Incubate **10 minutes** at **+70°C**.		
Binding preparation	Add **200 µl** of **ethanol 100%**. Homogenise the mixture by pipeting (~10 times) or by vortexing (~15 secondes).		
Transfer to columns and binding to the membrane	Identify columns, apply the **whole** obtained solution to the corresponding column. Centrifuge at 10 000 g/1 minute. *If the whole sample has not been loaded once, apply the residual volume onto the column and centrifuge 1 minute at 10 000 g.*		
1st wash	Change the collection tube and add **500 µl** of **BW buffer** to the column. Centrifuge 1 minute at 10 000 g.		
2nd wash	Change the collection tube and add **600 µl** of **B5 buffer** to the column. Centrifuge 1 minute at 10 000 g.		
Column dry step	Change the collection tube. Centrifuge 3 minutes at 10 000 g.		
Elution	Transfer the column to a microtube. Add **200 µl** of **BE buffer**. Incubate ~1 minute at room temperature and centrifuge 1 minute at 10 000 g.		
Storage	Close the tubes, identify and store at +2/8°C for 24 hours, then at <-15°C.		

*it's possible to centrifuge until 50 ml of water. In this case, after the discarding the supernatant is removed, add 1 ml of 1X PBS to the pellet, homogeneize and transfer 200 µl in the new microtube. And then, continue the protocol.

3. Extraction using ADIAMAG kit

See the NEKF user manual available of the web site mentioned on the certificate of analysis included in the used ADIAVET™ kit.

V. Amplification

a - Determine the number of analysed samples including the controls (e.g. positive and negative extraction controls, positive control of amplification (CTL+) and No Template Control (NTC)).

b - Defrost the A5 solution at room temperature. Vortex. Dispense **20 µl** of the A5 solution in each PCR tubes or PCR plate wells with a micropipette with a Nuclease-free tip.

c- **Immediately replace the A5 tube at <-15°C and in darkness.**

d- For each sample, the extraction negative control (obligatory) and the extraction positive control (recommended) add **5 µl** of purified extract to the 20 µl of A5 solution.
For the CTL+, add **5 µl** of the solution obtained in § II-3 to the 20 µl of A5 solution.
For the No Template Control (NTC), nothing is added to the A5 solution.
Immediately replace purified DNA extracts at +2/8°C or at **<-15°C**. Take care to have no bubbles in the bottom of the wells.

e- Once all the tubes have been prepared, run the real-time PCR amplification.

The *Leptospira* target is read in FAM. The Internal Control is read in VIC or HEX. The Quencher is non fluorescent. The solution contains a passive reference ROX for the ABI machines. Fluorescence is read during the elongation step (1 minute at 60°C).

The following program is defined for **ABI Prism** thermalcyclers (like 7500, StepOne…) from **Applied Biosystems** (check the "emulation 9600" option if available), for the **Rotorgene** from **Qiagen** and for the **Chromo 4** from **Biorad**:
2 minutes 50°C
10 minutes 95°C
15 seconds at 95°C and 1 minute at 60°C during 45 cycles

This program is concerning the **MX3000P** and **MX3005P** of **Stratagene**:
2 minutes 50°C
10 minutes 95°C
30 seconds at 95°C and 1 minute at 60°C during 45 cycles

Roche diagnostic: LightCycler 2*, LightCycler 480*
**** NOTE:*** *The use of LightCycler thermalcyclers requires a calibration manipulation. Adiagene will furnish process chart and reagents required for this calibration.*

Contact us if you wish to use other thermalcyclers.

VI. Interpretation of results

1. Definitions

The **« base line »** corresponds to the background of fluorescence and qualifies the non characteristic part of the curve observed during the first cycles.

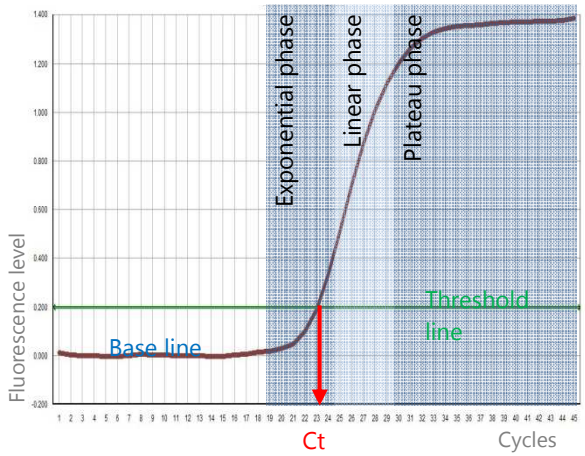
The **« Characteristic amplification curve »** qualifies a fluorescence curve with an exponential phase, a linear phase and a plateau phase.

The **« threshold line »** has to be placed over the background in the exponential phase of a characteristic amplification curve or a group of characteristic amplification curves.

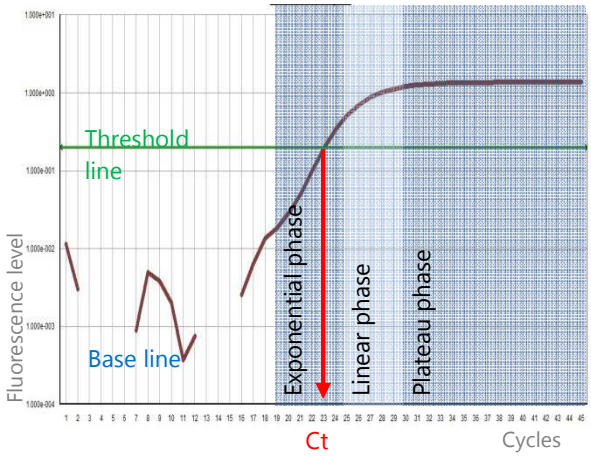
The **« threshold cycle » (Ct)** of a well corresponds, for each detected fluorophore, at the crossing point of the threshold line with the fluorescent curve. The Ct value expressed by the machine for each well depends on the threshold position and on the quantity of target sequences initially present in the PCR well.

Example of characteristic amplification curve

Arithmetic scale of Y axis



Logarithmic scale of Y axis



2. Validation and interpretation of results

Display the FAM curves from the plate and set the threshold value as indicated above.
Proceed in the same mean for the VIC/HEX curves.

A. Validation of the run

Amplification is **valid** if the following results are obtained for the controls:

Controls	No Template Control (NTC)	Amplification positive control (CTL+)	Extraction negative control	Extraction positive control *
FAM amplification	No	yes	no	yes
VIC/HEX amplification	yes	yes	yes	no/yes
Validation of	Absence of contamination for amplification	Amplification of the target	Absence of contamination for extraction	Extraction and amplification steps

* Optional

The indicative Ct values (FAM and VIC/HEX dyes) of the positive control (CTL+) are indicated in the certificate of analysis of the kit.

B. Result interpretation

DNA extraction and amplification for each sample are **valid** if at least a characteristic amplification curve is observed for *Leptospira* (FAM) or for the internal control (VIC/HEX).

Example	A	B	C	D
FAM amplification	no	yes	yes	no
VIC/HEX amplification	yes	no	yes	no
Result	negative	positive	positive	undetermined

The sample is considered as **negative** if a characteristic amplification curve is observed in VIC or HEX without any amplification in FAM (example A).

The sample is considered as **positive** if a characteristic amplification curve is observed in FAM (example B). Internal control can be co-amplified (example C).

A total absence of characteristic amplification curve for a sample (example D) shows a defective DNA extraction (lost or destruction of DNA) or a deficient real-time PCR (inhibitors in the sample, program error or no template added). In this case, we recommend first to repeat the test with pure and tenfold diluted DNA in Nuclease-free water. Then, if the test is still not valid, a new extraction is recommended.

VII. Index of symbols

Symbol	Meaning
	Catalogue number
	Manufacturer
	Upper temperature limit
	Use by date
	Batch code
	Consult Instructions for Use
	Contains sufficient for <n> tests
	Keep away from sunlight
	For veterinary in vitro use only – For animal use only

S.A.R.L. ADIAGENE
9, rue Gabriel Calloët-Kerbrat
22440 Ploufragan - France

RCS 417 876 299
Tel. +33 (0)2 96 68 40 20
www.biox.com