



ADIAVET™ MM/MI FAST TIME

TEST FOR THE DETECTION OF
MYCOPLASMA MELEAGRIDIS AND *MYCOPLASMA IOWAE*
BY REAL-TIME ENZYMATIC GENE AMPLIFICATION (PCR TEST)

References:

ADI601-100 (100 reactions)

ADI601-500 (500 reactions)



ADIAVET™ MM/MI FAST TIME

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Revision historic

| | |
|------------------|---|
| N/A | Not Applicable (first publication) |
| Correction | Correction of document anomalies |
| Technical change | Addition, revision and/or removal of information related to the product |
| Administrative | Implementation of non-technical changes noticeable to the user |

Note: minor typographical, grammar and formatting changes are not included in the revision historic

| Release Date | Part Number | Change type | Change summary |
|--------------|-------------|-------------|-------------------|
| 2020/01 | NE601-01 | N/A | First publication |

I. General informations

1. Purpose of the test

ADIAVET™ MM/MI FAST TIME kit is intended to detect *Mycoplasma meleagridis* and *Mycoplasma iowae* using real-time Polymerase Chain Reaction (PCR) technology from swab of poultry, environmental specimen, bacterial culture and FTA card.

2. Description and purpose of the test

This test is based on enzymatic gene amplification or PCR technology.

Amplified products are detected in real-time thanks to a specific labelled hydrolysis probe (5'-exonuclease technology).

The ADIAVET™ MM/MI FAST TIME kit enables the simultaneous detection of:

- *M. meleagridis* (probe labelled in FAM),
- *M. iowae* (probe labelled in Cy5),
- an internal control of amplification step specific from an exogenous DNA (probe labelled with a fluorochrome read in the same spectra as VIC and HEX).

Adiagene recommends using this test with DNA purification kits (Bio-X diagnostics, Qiagen). Other purification kits can be used if they have been validated by the user.

Analysis options according to the specimen:

| Specimen | Individual analysis | Pool of sample is possible*, up to |
|---|-------------------------------------|-------------------------------------|
| Swab on live animals (palate slit, tracheal...) | <input checked="" type="checkbox"/> | 6 |
| Swab on dead animals (joint, injured organ...) | <input checked="" type="checkbox"/> | 6 |
| Environmental specimen | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> |
| Bacterial culture (solid, liquid) | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> |
| FTA card | <input checked="" type="checkbox"/> | <input checked="" type="checkbox"/> |

* It depends on the epidemiological case and on the quality of the specimen.

II. Material & reagents

1. Reagents provided with the kit

| | | |
|---|---|--|
| REF ADI601-100 | | |
| A5 | Amplification Solution | 1 x 1000 µl tube green caps (ready to use) |
| MM CTL+ | Positive control <i>M. meleagridis</i> | 1 tube purple caps (dehydrated) |
| MI CTL+ | Positive control <i>M. iowae</i> | 1 tube purple caps (dehydrated) |
| EPC-Amp | exogenous internal control of amplification | 1 x 150 µl tube white caps (ready to use) |
| NF-Water | Nuclease free Water | 1 x 1000 µl tube with white cap (a ready-to-use reagent) |
| REF ADI601-500 | | |
| A5 | Amplification Solution | 5 x 1000 µl tubes green caps (ready to use) |
| MM CTL+ | Positive control <i>M. meleagridis</i> | 1 tube purple caps (dehydrated) |
| MI CTL+ | Positive control <i>M. iowae</i> | 1 tube purple caps (dehydrated) |
| EPC-Amp | exogenous internal control of amplification | 2 x 150 µl tubes white caps (ready to use) |
| NF-Water | Nuclease free Water | 1 x 1000 µl tube with white cap (a ready-to-use reagent) |
| Package insert downloadable from www.biox.com | | |

2. Validity and storage

On receipt, the kit should be stored at <-15°C.

It is recommended to make fractions of A5 solution if it should be defrosted more than 3 times.

Do not defrost reagents more than 3 times.

Realtime reagents are susceptible to light: store them in the darkness.

The A5 reagent is ready to use for PCR reaction.

Do not mix reagents of two different batches.

3. Use of MM CTL+

MM CTL+ is a positive control of amplification.

Add **200 µl** of **NF-Water** to the **MM CTL+** tube and vortex at least 20 seconds.

Aliquot this solution by 6 or 12 µl and store them to <-15°C.

For each analysis, use **5 µl** of **MM CTL+** in one of the wells.

4. Use of MI CTL+

MI CTL+ is a positive control of amplification.

Add **200 µl** of **NF-Water** to the **MI CTL+** tube and mix by vortexing at least 20 seconds.

Aliquot this solution by 6 or 12 µl and store them to <-15°C.

For each analysis, use **5 µl** of **MI CTL+** in one of the wells.

5. Used of EPC-Amp

The EPC-Amp should only be used if an extraction method other than ADIAPURE™ SLB is used, for example after extraction by silica column extraction or ADIAMAG.

Aliquot this solution and store it to <-15°C. **Do not defreeze each aliquot more than 3 times.**

For each PCR reaction, mix **0.5 µl** of **EPC-Amp** and **10 µl** of **A5 solution**.

6. Equipment required but not supplied in the kit

Material should be Nuclease-free (e.g. autoclaved 25 minutes twice at +120°C or once 60 minutes at +121°C)

- Thermal cycler with consumables for real-time PCR: 0.2 ml PCR tubes or closed 96-wells PCR plates with optical quality.
- Class II Microbiological Safety Cabinet
- Centrifuge for microtubes
- Etuve, heating baths or block heaters
- Vortex

- 1 - 10 µl pipette, 20 - 200 µl pipette and 200 - 1000 µl pipette
- Nuclease-free filter tips
- Nuclease-free microtubes: 1.5 ml and 2 ml
- Sterile tube of 5, 10 or 15 ml
- Latex or nitrile powder-free gloves
- 96-100% ethanol solution
- Nuclease-free water
- 2% Peptone water
- Sterile saline water (NaCl 8.5 g/l)

- DNA Extraction without purification:

- ADIAPURE™ SLB (Bio-X Diagnostics; 500 ml: ref. ADIADP01S1-500)
- ADIAPURE™ SLB (Bio-X Diagnostics; 100 ml: ref. ADIADP01S1-100)

or

- DNA extraction kit (individual silica columns)

- QIAamp® DNA Mini Kit (Qiagen, 50 tests: ref. 51304 or 250 tests: ref. 51306)

or

- Automated DNA/RNA extraction kit using magnetic beads

- ADIAMAG (Bio-X Diagnostics, 200 tests: ref. NADI003)

III. Recommendations before the analysis of samples

Before starting the test, read the entire protocol and scrupulously respect it.

1. Precautions

Adiogene has elaborated this PCR test with the use of Bio-X Diagnostics, Qiagen extraction kits. Other extraction kits can be used with a previous validation.

Follow the supplier's instructions for the storage, the preparation and the use of the extraction reagents.

Some kits include and/or need the use of toxic reagents. These reagents should be handled with gloves and into a chemical cabinet.

We strongly recommend that only appropriately trained personnel perform this test. Ensure the accuracy and precision of the micropipettes used. The quality of the obtained results depends upon rigorous respect of good laboratory practices.

The PCR generates large amount of amplified DNA. A few molecules of amplified products are sufficient to generate a positive result. It is important to reserve 2 rooms, one for manipulation of the samples to be tested, and another one for the amplified products analysis. Do not open the PCR tubes after amplification.

Samples for analysis should be handled and disposed of as biological waste. **Take all measures of security and confinement required for the manipulation of the concerned biological agents.**

We recommend using fractions of demineralised and saline water and to autoclave them 25 minutes at +120°C. Take a new fraction for each new manipulation to avoid contamination.

2. Storage of samples and DNA extracts

Samples can be stored a couple of days at +2/8°C. After 2 days, we recommend to store them at <-15°C.

Extracted DNAs are quite sensitive molecules. Crude extracts should be stored at the end of extraction at +2/8°C for 24 hours, then at <-15°C.

3. Controls preparation

The use of controls allows to verify the reliability of the results.

The controls are included per trial of analysis. A trial is defined as all the samples treated in the same conditions.

The amplification step, for all the types of samples, is validated with the association of the controls included in the kit.

- The exogenous internal control of amplification (EPC-Amp) added in A5 reagent verifies the amplification steps of each sample. (if using an extraction kit other than the ADIAPURE SLB kit)
- The exogenous internal control of extraction (included in the L3 buffer in ADIAPURE SLB extraction kit) added in all samples verifies the extraction and amplification steps of each sample.
- The CTL+ validate the amplification of the both targets.

Other controls must or could be added:

- **Negative control of extraction (required)**

To verify the absence of cross-contamination, at least one negative control must be included per trial. The control could be a negative sample or a buffer used for dilutions (sterile saline water or PBS 1X for example).

- **Positive control of extraction (recommended)**

A positive control could be added in each trial. The control is a sample including *M. meleagridis* and/or *M. iowae*. It could come from a positive sample available in the laboratory or from a negative sample spiked with a solution of *M. meleagridis* and/or *M. iowae*. This positive control should be closed to the limit of detection of the method. It will inform about the fidelity of the obtained results between different trials

IV. Extractions et purifications

1. Extraction with ADIAPURE™ SLB kit

See the user manual available of the web site mentioned on the certificate of analysis included in the used kit.

2. Extraction with ADIAMAG kit

See the user manual available of the web site mentioned on the certificate of analysis included in the used kit.

3. Extraction using QIAamp® DNA Mini kit

a) Preparation of samples

1. Swabs

Note: It is best to treat heavily soiled autopsy swabs individually.

1st method

Put **1 swab** or vortex successively **up to 3 swabs** in a microtube containing **180 µl** of **ATL buffer** and **20 µl** of **proteinase K**.

NB: if the swabs haven't been dipped, they may absorb all the solution. In this case, the amount of lysis buffer should be doubled.

Mix by vortexing

Press the swab against the rim of the microtube to collect as much buffer as possible.

See § IV 3b for the extraction and purification of DNA.

2nd method

Cut **1 to 3 swabs** in a tube (e.g. a 5 ml – tube) containing **700 µl** of **saline water**.

Vortex ~10 seconds.

Transfer **200 µl** of **supernatant** in a microtube containing **180 µl** of **ATL buffer** and **20 µl** of **proteinase K**.

Mix by vortexing

See § IV 3b for the extraction and purification of DNA.

NB: swabs in saline water can be stored at +2/8°C.

2. From solid culture

Scrape the medium with a swab (dipped in 2% peptone water).

Put it in a microtube containing **180 µl** of **ATL buffer** and **20 µl** of **proteinase K**.

Vortexing ~5 seconds.

Press the swab against the rim of the microtube to collect as much buffer as possible.

See § IV 3b for the extraction and purification of DNA.

3. From liquid culture

Centrifuge (20 minutes at 10 000 g) **100 to 500 µl** of **culture** in a microtube

Discard the supernatant.

Add **180 µl** of **ATL buffer** and **20 µl** of **proteinase K**.

Mix by vortexing

See § IV 3b for the extraction and purification of DNA.

4. From drag swabs

Add **30 ml** of **peptone water** in the bag containing the drag swab.

Homogenize by mixing the bag.

NB: If the drag swab has absorbed all the liquid, add 10 ml more peptone water and mix the bag again.

Transfer **200 µl** of **supernatant** in a microtube containing **180 µl** of **ATL buffer** and **20 µl** of **proteinase K**. Mix by vortexing

See § IV 3b for the extraction and purification of DNA.

5. From FTA card

Cut the 1 cm² of FTA card and introduce its in a microtube

Add **1 ml** of saline water

Incube overnight at room temperature

Transfer **200 µl** of supernatant in a microtube containing **180 µl** of **ATL buffer** and **20 µl** of **proteinase K**. Mix by vortexing

See § IV 3b for the extraction and purification of DNA.

b) protocol

All the centrifugations are performed at room temperature.

| | Swab / Bacterial culture / drag swab / FTA card |
|---|--|
| Preparation of the sample | See § IV.3.a |
| Lysis | Incubate 15 minutes at +56°C . |
| | Add 200 µl of AL buffer . Vortex. Incubate 10 minutes at +70°C . |
| Binding preparation | Add 200 µl of ethanol 100% . Homogenise the mixture by pipeting (~10 times) or by vortex (~15 secondes). |
| Transfer to columns and binding to the membrane | Identify columns, apply the whole obtained solution to the corresponding column. Centrifuge 1 minute at 10 000 g. <i>If the whole sample has not been loaded once, apply the residual volume onto the column and centrifuge 1 minute at 10 000 g.</i> |
| 1 st wash | Change the collection tube and add 500 µl of AW1 buffer to the column. Centrifuge 1 minute at 10 000 g. |
| 2 nd wash | Change the collection tube and add 500 µl of AW2 buffer to the column. Centrifuge 1 minute at 10 000 g. |
| Column dry step | Change the collection tube. Centrifuge 3 minutes at 10 000 g. |
| Elution | Transfer the column to a microtube. Add 200 µl of AE buffer . Incubate ~1 minute at room temperature and centrifuge 1 minute at 10 000 g. |
| Storage | Close the tubes, identify and store at +2/8°C for 24 hours, then at <-15°C. |

V. Amplification

a - Determine the n number of analysed samples including the controls (e.g. positive and negative extraction controls, positive control of amplification (CTL+) and No Template Control (NTC)).

b - Defrost the A5 solution at room temperature. Vortex and prepare the amplification solution A5:

Extraction with ADIAPURE™ SLB:

Place **10 µl** of A5 solution in each PCR tubes or PCR plate wells.

Other extraction:

Place $(n+1)*10 \mu\text{l}$ of A5 solution in microtube,

Add $(n+1)*0,5 \mu\text{l}$ « EPC-Amp ».

Distribute **10 µl** of the solution in each PCR tubes or PCR plate wells

c- **Immediately replace the A5 tube at <-15°C and in darkness.**

d- For each sample, the extraction negative control (obligatory) and the extraction positive control (recommended) add **5 µl** of purified extract to the 10 µl of A5 solution.

For the CTL+, add **5 µl** of the solution obtained in § II-3 or § II-4 to the 10 µl of A5 solution.

For the No Template Control (NTC), nothing is added to the A5 solution.

Immediately replace purified DNA extracts at +2/8°C or at <-15°C. Take care to have no bubbles in the bottom of the wells.

e- Once all the tubes have been prepared, run the real-time PCR amplification as soon as possible.

The *M. meleagridis* target is read in FAM. The *M. iowea* target is read in Cy5. The Internal Control is read in VIC or HEX. The Quencher is non-fluorescent. The solution contains a passive reference ROX for the ABI machines. Fluorescence is read during the elongation step (at 60°C).

The following programs are defined for **ABI Prism** thermalcyclers (like 7500, StepOne, QS5...) from **Applied Biosystems** (check the "emulation 9600" option if available), for the **MX3005P** and **ARIAMX** of **Agilent** and for **CFX96** of **BioRad**.

| Standard Program | | FAST program | |
|---------------------------|-----------|--------------|-----------|
| 2 min 50°C 10 min 95°C | | 2 min. 95°C | |
| 15 sec | 45 cycles | 5 sec 95°C | 45 cycles |
| 1 min | | 30 sec 60°C | |

Contact us if you wish to use other thermalcyclers.

VI. Interpretation of results

1. Definitions

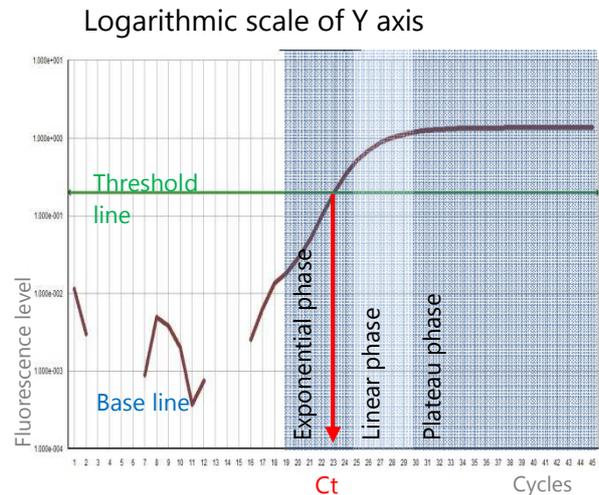
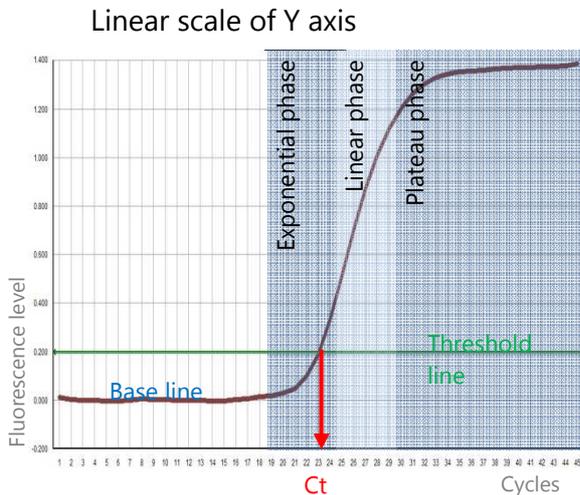
The « **base line** » corresponds to the background of fluorescence and qualifies the non-characteristic part of the curve observed during the first cycles.

The « **Characteristic amplification curve** » qualifies a fluorescence curve with an exponential phase, a linear phase and a plateau phase.

The « **threshold line** » has to be placed over the background in the exponential phase of a characteristic amplification curve or a group of characteristic amplification curves.

The « **threshold cycle** » (**Ct**) of a well corresponds, for each detected fluorophore, at the crossing point of the threshold line with the fluorescent curve. The Ct value expressed by the machine for each well depends on the threshold position and on the quantity of target sequences initially present in the PCR well.

Example of characteristic amplification curve:



2. Validation and interpretation of results

*Display the FAM curves from the plate and set the threshold value as indicated above.
Proceed in the same mean for the VIC/HEX and Cy5 curves.*

a. Validation of the run

Amplification is **valid** if the following results are obtained for the controls:

| Controls | | No Template Control (NTC) | Amplification positive control (MM CTL+) | Amplification positive control (MI CTL+) | Extraction negative control | Extraction positive control * |
|-----------------------|---|--|---|--|---|------------------------------------|
| FAM amplification | | no | yes | no | no | yes |
| Cy5 amplification | | no | no | yes | no | yes |
| Amplification VIC/HEX | If ADIAPURE SLB extraction (EPC-Amp not added in mix) | no | no | no | yes | yes |
| | If other extractions (EPC-Amp added in mix) | yes | yes | yes | yes | yes |
| Validation of | | Absence of contamination for amplification | Amplification of <i>M. meleagridis</i> target | Amplification of <i>M. iowaea</i> target | Absence of contamination for extraction | Extraction and amplification steps |

* Optional

The indicative Ct values (FAM, Cy5 and VIC/HEX dyes) of the CTL+ are indicated in the certificate of analysis of the kit.

b. Result interpretation

DNA extraction and amplification for each sample are **valid** if at least a characteristic amplification curve is observed for *M. meleagridis* (FAM), for *M. iowaea* (Cy5) or for the internal control (VIC/HEX).

| Example | A | B | C | D | E |
|-----------------------|----------|------------------------------------|-------------------------------|---|--------------|
| FAM amplification | No | Yes | No | Yes | No |
| Cy5 amplification | No | No | Yes | Yes | No |
| VIC/HEX amplification | Yes | Yes/No | Yes/No | Yes/No | No |
| Results | Negative | Positive for <i>M. meleagridis</i> | Positive for <i>M. iowaea</i> | Positive for <i>M. meleagridis</i> and <i>M. iowaea</i> | Undetermined |

The sample is considered as **No detected** if a characteristic amplification curve is observed in VIC or HEX without any amplification in FAM (example A).

The sample is considered as **Detected** if a characteristic amplification curve is observed in FAM (examples B and D) and/or in Cy5 (examples C and D). Internal control can be co-amplified.

A total absence of characteristic amplification curve for a sample (example E) shows a defective DNA extraction (loss or destruction of DNA) or a deficient real-time PCR (inhibitors in the sample, program error or no template added). In this case, we recommend first to repeat the test with pure and tenfold diluted DNA in Nuclease-free water. Then, if the test is still not valid, a new extraction is recommended.

VII. Index of symbols

| Symbol | Meaning |
|--|--|
|  | Catalogue number |
|  | Manufacturer |
|  | Upper temperature limit |
|  | Use by date |
|  | Batch code |
|  | Consult Instructions for Use |
|  | Contains sufficient for <n> tests |
|  | Keep away from sunlight |
|  | For veterinary in vitro use only – For animal use only |

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**S.A.R.L. ADIAGENE**
9, rue Gabriel Calloët-Kerbrat
22440 Ploufragan - France

RCS 417 876 299
Tel. +33 (0)2 96 68 40 20
www.biox.com