



Instruction manual ADL02Y1\_LAWI\_NO\_(EN)\_V01 01/2025

# **LAWSONIA**

Reference: ADLO2Y1-100

Test for the detection of Lawsonia intracellularis by real time enzymatic amplification

PCR Test - 100 reactions

For veterinary in vitro use only



Sample	Individual analysis	Pool of sample possible*, up to:
Swab (rectal)	✓	3
Tissue (intestinal mucosa, etc.)	✓	*
Faeces	✓	3
Environmental sample	✓	×
Oral fluid	✓	*

<sup>\*</sup> Depending on the epidemiological case and on the quality of samples

# Kit composition

Content		ADL02Y1-100 Kit
	Comem	100 reactions
A6	Amplification solution	1 lyophilized vial with blank caps
Ab		(To reconstitute)
Debudenties buffer	Rehydration solution	1 x 6 mL vial
Rehydration buffer		(Ready to use)
LAW CTL+	1	1 tube with purple cap
LAW CTL+	Lawsonia intracellularis positive control	(To reconstitute)
NF-Water	NE Meter	1 x 1000 μL tube with white cap
NF-Water Nuclease-Free Water		(Ready to use)

# **Revision history**

Date	Version	Modifications
01/2025	V01	First version

Note: minor typographical, grammar and formatting changes are not included in the revision history.

### A. Introduction

Lawsonia intracellularis is the ethological agent of Proliferative Enteropathy (PE), causing a disease commonly known as 'lleitis' and having a major economic impact on the pig industry. L. intracellularis has two clinical presentations in pigs:

Porcine hemorrhagic enteropathy (PHE) is the acute, haemorrhagic and severe form, affecting animals aged between 4 and 12 months. Porcine intestinal adenomastosis (PIA) is the most common form. It is a chronic infection diagnosed in pigs aged between 6 and 20 weeks. Infection by *L. intracellularis* is spread mainly via the faecal-oral route. Infected animals excrete the bacteria in their feces, and contamination often occurs in confined conditions, such as in intensive livestock farms.

### B. Test principle

ADIALYO™ LAWSONIA test is based on the amplification of specific *Lawsonia intracellularis* DNA. This test is intended to detect simultaneously, in one well:

- L. intracellularis (FAM labelled probe).
- Internal control of amplification specific from an exogenous DNA (HEX labelled probe or its equivalent).

# C. Storage conditions

- Store the kit at a temperature below +2/8 °C after reception.
- Store away from sunlight and keep dry.
- After reconstitution, prepare aliquots and store them at a temperature below -15 °C until the expiration date.
- Do not thaw more than 3 times.

### D. Material required but not provided

- Real-time Thermal cycler and device.
- Instrument for homogenous mixing of tubes.
- Pipettes of 1 10 μL, 20 200 μL and 200 1000 μL.
- Nuclease-free filtered pipette tips.
- Nuclease-free microtubes of 1,5 mL and 2 mL.
- Powdered-free latex or nitrile gloves.
- Nuclease-free water.
- Kit for nucleic acids extraction.

Additional kits (on request) for method adoption and PCR

LD<sub>PCR</sub> Positive Control – Law (Ref.: ADC02LD)

Confirmation of performances – LOD<sub>PCR</sub> of kit.

# E. Warnings and precautions

- For veterinary in vitro use only.
- For animal use only.
- For professional use only.
- All instructions must be read before performing the test and strictly respected.
- Do not use reagents after the expiration date.
- Do not use reagents if the packaging is damaged.
- Do not open PCR wells or tubes after amplification.
- Do not mix reagents from different batches.
- Used material must be disposed of in compliance with the legislation in force regarding environmental protection and biological waste management.
- This kit contains products of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not totally guarantee the absence of transmissible pathogenic agents. It is therefore recommended that these products be treated as potentially infectious and handled observing the usual safety precautions (do not ingest or inhale).

### F. Nucleic acids extraction

#### 1. Extraction kits

Nucleic acids must be extracted from the samples before using the kit. The RNA/DNA purification kits listed below are recommended by Bio-X Diagnostics:

Product name	Extraction system	Number of tests and reference
ADIAMAG	Magnetic beads	200 tests: ref. NADI003 800 tests: ref. NADI003-XL
ADIAPURE Lysis Flex	Direct lysis	500 mL: ref. ADPLF1-500

For the extraction, consult the user manual version, available on the website, indicated on the certificate of analysis included in the PCR kit of interest.

Extraction protocols are described in validation data. Other purification kits can be used if they have been validated by the user.

After extraction, nucleic acid extracts can be kept on ice or at +2/8 °C for a few hours or until use. For long term storage, they must be kept at a temperature below -15 °C or -65 °C.

#### 2. Controls

Using controls allows to verify the reliability of the results.

Control	Validation of	Usage
No Template Control (NTC)	Absence of amplification contamination	5 µL NF-Water in a well per run
LAW CTL+	LAW target amplification	5 μL CTL+ in a well per run
Negative extraction control	Absence of contamination for the extraction and amplification	1 extraction (water or lysis buffer) per run
Positive extraction control	Extraction and amplification	1 extraction (Positive sample between 1 et 100X LOD <sub>METHOD</sub> ) per run

### G. Procedure

### 1. Amplification solution A6 preparation

- Add 1000 μL of « Rehydration buffer » per A6 tube.
- Homogenize tube contents using a mixer, such as vortex, at least 20 seconds.
- After reconstitution, aliquot the solution and store at a temperature below -15 °C until the expiration date. Do not thaw more than 3 times.
- To use the A6, please refer to § « Amplification », Step 1.

### 2. Preparation of CTL+ control

- Add 200 μL of « NF-Water » per tube.
- Homogenize the tubes by suction and pressure, then use a mixer, such as vortex, > 20 seconds and until dissolution of the blue pellet.
- After reconstitution, aliquot and store the solution at a temperature below -15 °C until the kit expiration date. Do not thaw more than 3 times.
- For each assay, use **5 μL** of CTL+ in one of the dedicated wells (see § « Amplification », Step 2).

# 3. Amplification

#### Warning:

- Before starting, rehydrate or thaw reagents at room temperature in the dark.
- Homogenize all reagents and samples before use.
- Store reagents at a temperature below -15 °C after use.

Step 1: Dispense 10 μL of amplification solution (A6) per well.

<u>Step 2:</u> Dispense 5  $\mu$ L of nucleic acids extracts and 5  $\mu$ L of controls in each dedicated well.

Use NF-Water for the No Template Control (NTC).

<u>Step 3:</u> Cover the wells with an appropriate optical film or caps. Centrifuge briefly (recommended).

Step 4: Set up the amplification program.

The following program is defined for ABI Prism thermocyclers (like 7500, QuantStudio5, Step-one...) from Applied Biosystems, for Mx3000, Mx3005P and AriaMx from Agilent, for LightCycler from Roche Diagnostics, for Rotor-Gene Q from Qiagen, for CFX96 and Chromo 4 from Biorad and for MIC from BioMolecular System.

DNA/RNA Program		
10 min. 45 °C		
2 min. 95 °C		
5 sec. 95 °C	40 avalos	
30 sec. 60 °C*	40 cycles	

<sup>\*</sup>Reading and parameters for fluorescence acquisition:

Fluorochrome	Absorbance (nm)	Emission (nm)
FAM	494	520
HEX or equivalent	530	549
ROX	575	602

**Note:** The Quencher is non-fluorescent. The A6 solution contains a passive reference read in the same spectra as ROX for ABI machines.

For other thermal cycler instruments, please contact your sales representative or the customer relations department.

### H. Reading and interpretation

Display all curves and position the threshold line for each fluorochrome.

### 1. Test validation

Amplification is valid if the following results are obtained. Expected Ct (Threshold Cycle) values for the CTL+ are indicated on the certificate of analysis of the kit.

	Amplification		
Controls	FAM	HEX or equivalent	Validation of
No Template Control (NTC)	No	Yes	Absence of amplification contamination
LAW CTL+	Yes	Yes	Target amplification
Extraction negative control	No	Yes	Absence of extraction contamination
Extraction positive control	Yes	Yes	Extraction and amplification steps

### 2. Results interpretation

Nucleic acids extraction and amplification are **valid** for each sample if at least one typical amplification curve is observed in FAM and/or HEX or equivalent.

Amplification		Interpretation
FAM	HEX or equivalent	Lawsonia intracellularis
No	Yes	Undetected
Yes	Yes	Detected
Yes	No	Detected
No	No	Undetermined

#### « Undetermined »: no characteristic amplification curve.

### Possible causes:

Defective PCR due to inhibitors, set up error, absence of samples, degraded samples and/or issue with nucleic acids extraction (loss or destruction of nucleic acids).

#### Recommendations:

Set up a new PCR assay using pure nucleic acids extracts and 10x dilutions in Nuclease-free water.

If the assay is inconclusive, perform a new nucleic acids extraction.

### **Symbols**

Symbole	Signification	
REF	Catalog number	
<b></b>	Manufacturer	
1	Temperature limitation	
	Use by	
LOT	Batch code	
Ti	Consult Instructions for Use	
$\sum$	Contain sufficient for "n" tests	
VET	For veterinary <i>in vitro</i> use only – For animal use only	
*	Keep away from sunlight	
Ţ	Keep dry	

Extract nucleic acids with



Adia or



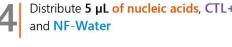
Scan me to discover Adiamag™

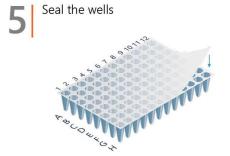
Add 1000 µL of Rehydration buffer to the A6 amplification solution

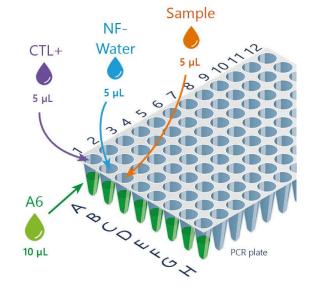


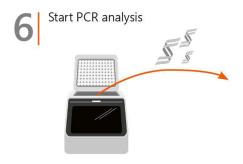
Distribute 10 µL of A6 amplification solution

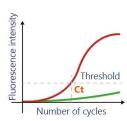
Distribute 5 µL of nucleic acids, CTL+ and NF-Water











\*The notes do not replace the instructions for use of which they are a summary.

**Contact us** 

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